

GBSC 724 Metabolomics Class

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Setting up for MS-DIAL

- **Enter “MS-DIAL” into your browser and download MS-DIAL vs 4.12**
- **Scroll down to find the MSMS library files**
 - For this exercise, download the public negative ion library and place on the desktop
- **Click on ABF converter tab (top of the home page)**
 - Click download – this will open a box – enter your name, institution, country and email address – an email will be sent to you
 - Open the email and download ABD converter – unzip the file
 - Open the ABF converter folder and scroll down to find the program and double click to open it

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Getting prepared for conversion to .ABF files

Name	Date modified	Type	Size
<input type="checkbox"/> Neg_C4.wiff	3/14/2017 1:54 PM	WIFF File	5,868 KB
<input type="checkbox"/> Neg_C4.wiff.scan	3/14/2017 1:54 PM	SCAN File	125,715 KB
<input type="checkbox"/> Neg_C5.wiff	3/14/2017 4:26 AM	WIFF File	6,068 KB
<input type="checkbox"/> Neg_C5.wiff.scan	3/14/2017 4:26 AM	SCAN File	118,180 KB
<input type="checkbox"/> Neg_C6.wiff	3/14/2017 6:02 PM	WIFF File	5,672 KB
<input type="checkbox"/> Neg_C6.wiff.scan	3/14/2017 6:02 PM	SCAN File	113,206 KB
<input type="checkbox"/> Neg_G4.wiff	3/14/2017 11:09 AM	WIFF File	6,136 KB
<input type="checkbox"/> Neg_G4.wiff.scan	3/14/2017 11:09 AM	SCAN File	129,339 KB
<input type="checkbox"/> Neg_G5.wiff	3/14/2017 3:17 PM	WIFF File	6,056 KB
<input type="checkbox"/> Neg_G5.wiff.scan	3/14/2017 3:17 PM	SCAN File	125,400 KB
<input type="checkbox"/> Neg_G6.wiff	3/14/2017 3:04 AM	WIFF File	6,092 KB
<input type="checkbox"/> Neg_G6.wiff.scan	3/14/2017 3:04 AM	SCAN File	114,121 KB

Open conversion box

3

Creating .abf files

Name	Date modified	Type	Size
<input type="checkbox"/> Neg_C4.abf	2/10/2020 4:42 PM	ABF File	609,340 KB
<input type="checkbox"/> Neg_C4.wiff	3/14/2017 1:54 PM	WIFF File	5,868 KB
<input type="checkbox"/> Neg_C4.wiff.scan	3/14/2017 1:54 PM	SCAN File	125,715 KB
<input type="checkbox"/> Neg_C5.abf	2/10/2020 4:42 PM	ABF File	574,528 KB
<input type="checkbox"/> Neg_C5.wiff	3/14/2017 4:26 AM	WIFF File	6,068 KB
<input type="checkbox"/> Neg_C5.wiff.scan	3/14/2017 4:26 AM	SCAN File	118,180 KB
<input type="checkbox"/> Neg_C6.abf	2/10/2020 4:43 PM	ABF File	544,464 KB
<input type="checkbox"/> Neg_C6.wiff	3/14/2017 6:02 PM	WIFF File	5,672 KB
<input type="checkbox"/> Neg_C6.wiff.scan	3/14/2017 6:02 PM	SCAN File	113,206 KB
<input type="checkbox"/> Neg_G4.abf	2/10/2020 4:43 PM	ABF File	624,544 KB
<input type="checkbox"/> Neg_G4.wiff	3/14/2017 11:09 AM	WIFF File	6,136 KB
<input type="checkbox"/> Neg_G4.wiff.scan	3/14/2017 11:09 AM	SCAN File	129,339 KB
<input type="checkbox"/> Neg_G5.abf	2/10/2020 4:44 PM	ABF File	607,292 KB
<input type="checkbox"/> Neg_G5.wiff	3/14/2017 3:17 PM	WIFF File	6,056 KB
<input type="checkbox"/> Neg_G5.wiff.scan	3/14/2017 3:17 PM	SCAN File	125,400 KB
<input type="checkbox"/> Neg_G6.abf	2/10/2020 4:44 PM	ABF File	549,515 KB
<input type="checkbox"/> Neg_G6.wiff	3/14/2017 3:04 AM	WIFF File	6,092 KB
<input type="checkbox"/> Neg_G6.wiff.scan	3/14/2017 3:04 AM	SCAN File	114,121 KB

Completed

Raw File Conversion is successfully completed.

OK

4

Loading MS-DIAL

Name	Date modified	Type	Size
MonaRestApi.dll	1/2/2020 12:28 AM	Application extens...	35 KB
MonaRestApi.dll.config	4/1/2019 11:03 PM	CONFIG File	2 KB
MSDIAL	1/7/2020 4:12 PM	Application	6,846 KB
MSDIAL.exe.config	9/20/2018 12:40 PM	CONFIG File	2 KB
MSDIAL	11/24/2019 4:38 PM	Configuration setti...	1 KB
MsdialCommon.dll	1/7/2020 4:12 PM	Application extens...	45 KB
MsdialConsoleApp	1/7/2020 4:12 PM	Application	132 KB
MsdialConsoleApp.exe.config	7/1/2016 12:39 PM	CONFIG File	1 KB
MsdialDataExporter.dll	5/30/2019 4:29 PM	Application extens...	41 KB
MsdialGcmsProcess.dll	1/7/2020 4:12 PM	Application extens...	165 KB

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The screenshot displays the MS-DIAL software interface with the following components:

- File navigator:** Shows a list of files and folders.
- Peak spot navigator:** Includes a label dropdown (set to 'None'), a peak spots slider (set to 100%), and a display filter section with checkboxes for 'Ref. matched', 'Suggested', 'CCS matched', 'Unknown', 'MS2 acquired', 'Molecular ion', 'Blank filter', and 'Unique ions'.
- EIC of focused spot:** A plot of relative abundance (0-100) versus retention time (0-100 min).
- Bar chart of aligned spot:** A plot showing relative abundance for aligned spots.
- EIC of aligned spot:** A plot showing relative abundance for aligned spots.
- Basic peak property / Compound detail:** Fields for Annotation, RT [min], CCS, m/z, Peak height [area], Formula [Ontology], InChIKey, and Comment.
- Survey scan (MS1) spectrum:** A plot of relative abundance (0-100) versus m/z (0-100).
- Peak spot viewer / Alignment spot viewer:** A plot of m/z (0-100) versus retention time (0-100 min).
- Measurement vs. Reference:** A plot comparing relative abundance (0-100) versus m/z (0-100) for a measurement and a reference.
- Alignment navigator:** A vertical panel on the left side.
- Region focus by ID:** Input fields for ID, RT (min), and m/z, each with a '-1' value and a search icon.
- Information box:** Located at the bottom right, displaying 'Ctr+D, metabolite name to 'Unknown''.

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Setting the initial parameters

Start up a project

Project file path: C:\Users\sbarne\Desktop\Class 2020\class 2-10-20 files\2020_2_11_7_32_59.mtd Select the path to the .abf files

Ionization type

Soft ionization (LC/MS, LC/MS/MS, or precursor-oriented GC/MS/MS)

Hard ionization (GC/MS)

Separation type

Chromatography (GC, LC, CE, or SFC)

Ion mobility (now coupled with liquid chromatography)

MS method type

Conventional LC/MS or data dependent MS/MS

SWATH-MS or conventional All-ions method All-ions with multiple CEs (cycled like 0V-10V-40V)

Experiment file:

Data type (MS1)

Profile data

Centroid data

Data type (MS/MS)

Profile data

Centroid data

Ion mode

Positive ion mode

Negative ion mode

Target omics

Metabolomics

Lipidomics

Advanced: add further meta data

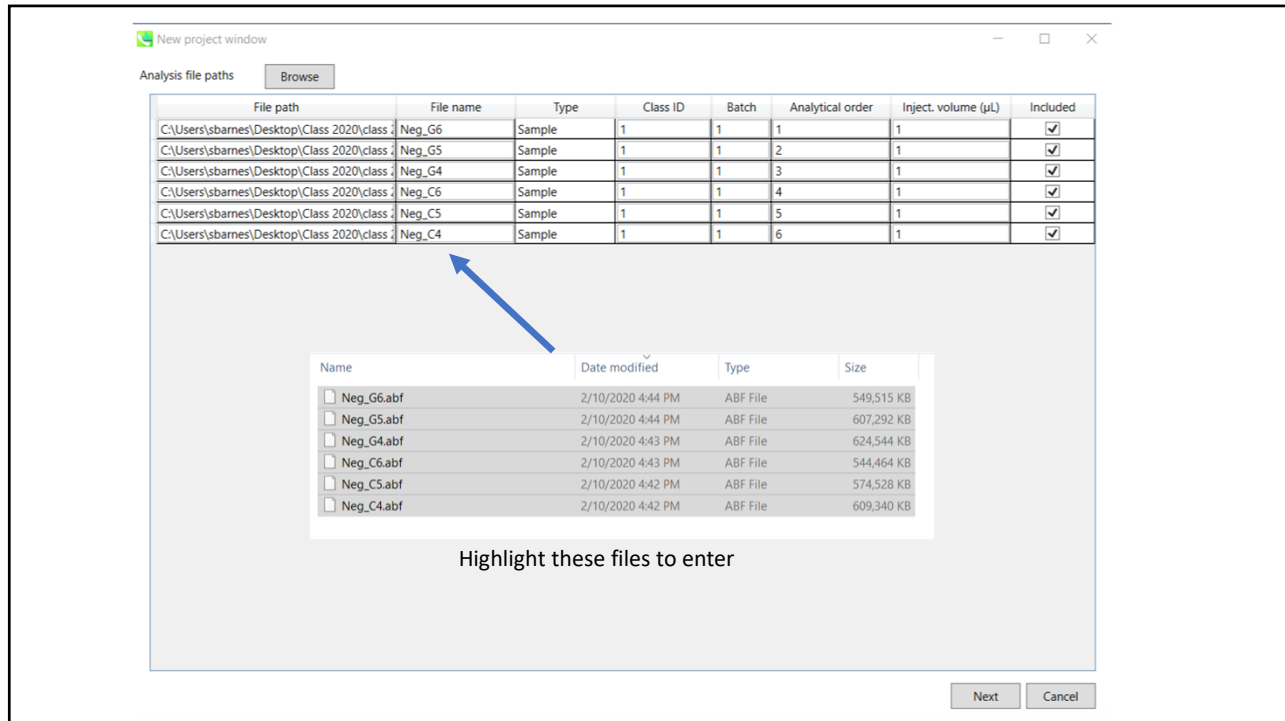
7

New project window

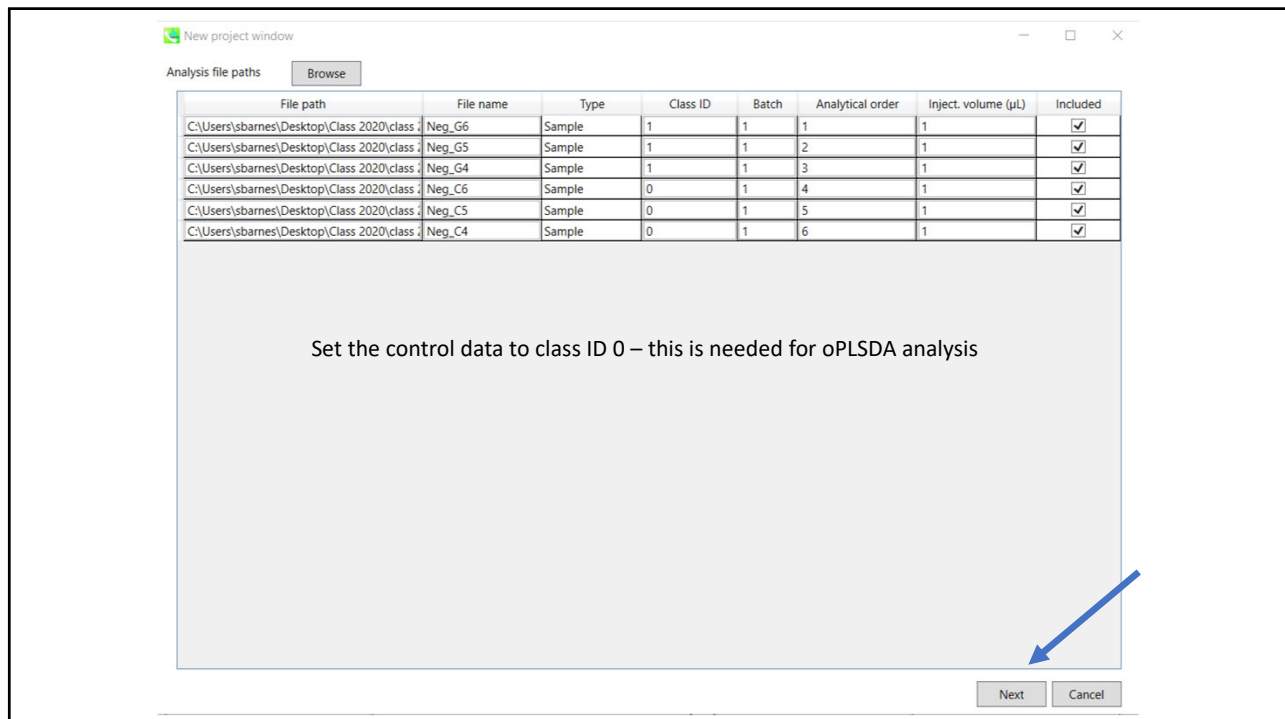
Analysis file paths Click to find the .abf files

File path	File name	Type	Class ID	Batch	Analytical order	Inject. volume (µL)	Included

8



9



10

Setting the parameters under the tabs

Analysis parameter setting

Data collection Peak detection MS2Dec Identification Adduct Alignment Mobility Isotope tracking

Mass accuracy

MS1 tolerance: Da

MS2 tolerance: Da

Advanced

Changed

Do not click the finish tab in the righthand corner until later

11

No changes to these tabs

Analysis parameter setting

Data collection Peak detection MS2Dec Identification Adduct Alignment Mobility Isotope tracking

Peak detection parameters

Minimum peak height: amplitude

Mass slice width: Da

Advanced

Analysis parameter setting

Data collection Peak detection MS2Dec Identification Adduct Alignment Mobility Isotope tracking

Deconvolution parameters

Sigma window value:

MS/MS abundance cut off: amplitude

Advanced

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Need to locate the data library

Analysis parameter setting

Data collection Peak detection MS2Dec Identification Adduct Alignment Mobility Isotope tracking

MSP file and MS/MS identification setting

MSP file: Select

Retention time tolerance: min

Accurate mass tolerance (MS1): Da

Accurate mass tolerance (MS2): Da

Identification score cut off: %

Use retention time for scoring:

Use retention time for filtering:

Advanced

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The database has been added

Analysis parameter setting

Data collection Peak detection MS2Dec Identification Adduct Alignment Mobility Isotope tracking

MSP file and MS/MS identification setting

MSP file: Select

Retention time tolerance: min

Accurate mass tolerance (MS1): Da

Accurate mass tolerance (MS2): Da

Identification score cut off: %

Use retention time for scoring:

Use retention time for filtering:

Advanced

This has been reset

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Check the boxes for possible adducts

Analysis parameter setting

Data collection Peak detection MS2Dec Identification Adduct Alignment Mobility Isotope tracking

Adduct ion setting User-defined adduct

Molecular species	Charge	Accurate mass [Da]	Included
[M-H]-	1	-1.00782503207	<input checked="" type="checkbox"/>
[M-H ₂ O-H]-	1	-19.01838971207	<input checked="" type="checkbox"/>
[M+Na-2H]-	1	20.97411921676	<input checked="" type="checkbox"/>
[M+Cl]-	1	34.96885268	<input type="checkbox"/>
[M+K-2H]-	1	36.94805661586	<input type="checkbox"/>
[M+FA-H]-	1	44.99765396793	<input checked="" type="checkbox"/>
[M+Hac-H]-	1	59.01330396793	<input type="checkbox"/>
[M+C ₂ H ₃ N+Na-2H]-	1	62.00066831777	<input type="checkbox"/>
[M+Br]-	1	78.9183371	<input type="checkbox"/>
[M+TFA-H]-	1	112.98503896793	<input type="checkbox"/>
[M-C ₆ H ₁₀ O ₄ -H]-	1	-147.06573383101	<input type="checkbox"/>
[M-C ₆ H ₁₀ O ₅ -H]-	1	-163.06064845057	<input type="checkbox"/>
[M-C ₆ H ₈ O ₆ -H]-	1	-177.03991300599	<input type="checkbox"/>
[M+CH ₃ COONa-H]-	1	80.99524996793	<input type="checkbox"/>
[2M-H]-	1	-1.00782503207	<input checked="" type="checkbox"/>
[2M+FA-H]-	1	44.99765396793	<input checked="" type="checkbox"/>
[2M+Hac-H]-	1	59.01330396793	<input type="checkbox"/>
[3M-H]-	1	-1.00782503207	<input checked="" type="checkbox"/>
[M-2H] ²⁻	2	-2.01565006414	<input checked="" type="checkbox"/>
[M-3H] ³⁻	3	-3.02347509621	<input checked="" type="checkbox"/>

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Analysis parameter setting

Data collection Peak detection MS2Dec Identification Adduct Alignment Mobility Isotope tracking

Alignment parameters setting

Result name: alignmentResult_2020_2_11_7_39_25

Reference file: Neg_G6

Retention time tolerance: 1 min

MS1 tolerance: 0.015 Da

Advanced

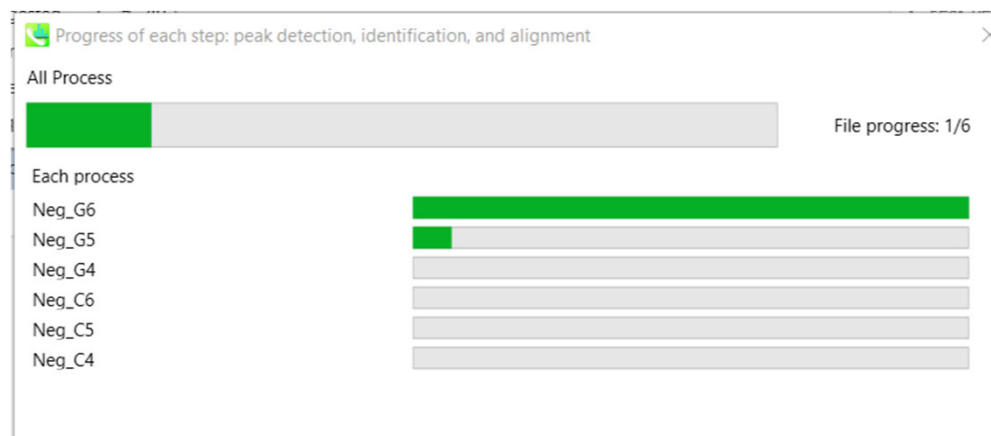
Now you can press Finish

Load Together with Alignment Finish Cancel

This time has been reset

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Showing the progress of the analysis



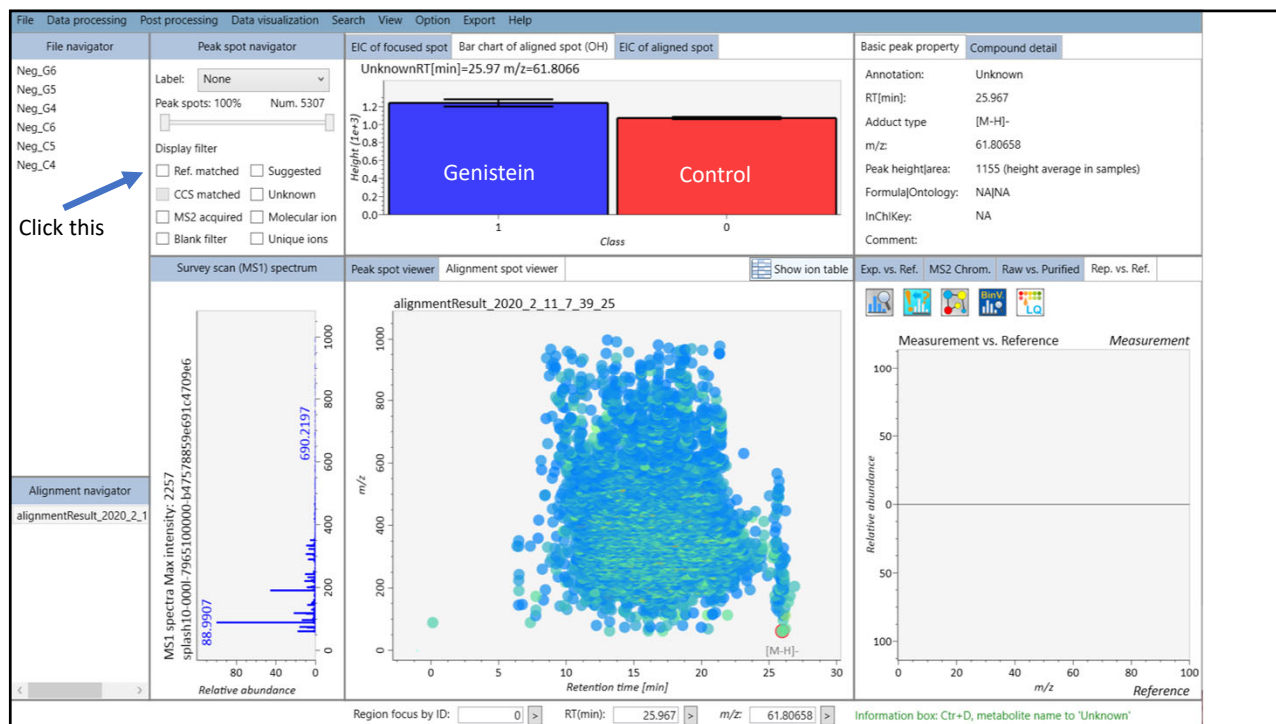
17

When the green bar is fully over to the right, secondary programs will run. Finally, the MS-DIAL analysis panes will appear.

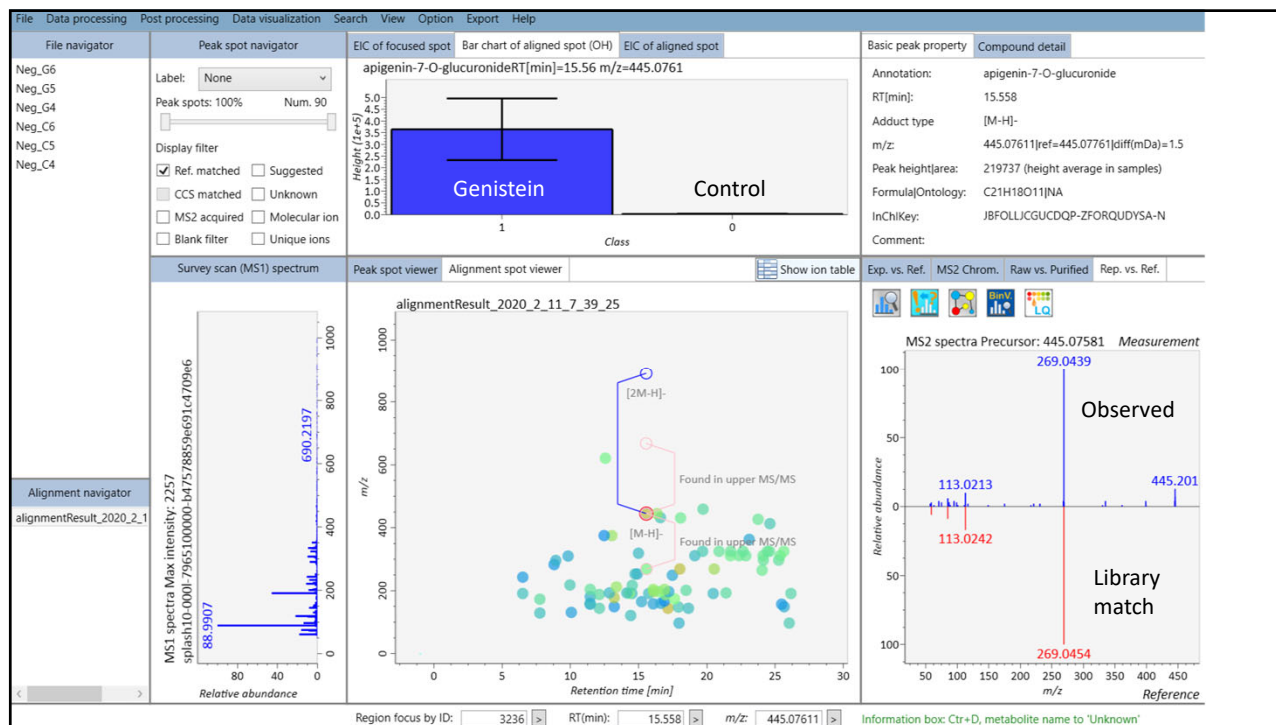
Double click on the alignment result in the left side bar

All the detected ions (known and unknown) are displayed

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Click on "Ion table" to show the "identified" ions

Alignment Table

Num of rows: 81

Metabolite Name Filter:

Comment Filter:

61.81 Mz Range 997.29 0.1 RT Range 26.9

ID	RT(min)	m/z	Type	Fill %	Metabolite name	Comment	Correlation	S/N	ANOVA P-value	Fold change (Max/Min)	BarChart
1702	24.43	325.1866	[M-H]-	0.33	Dodecylbenzenesulfonic		-0.57	754.0	4.66E-01	1.05	
2187	23.10	362.9694	[M-H]-	0.83	Perfluoroheptanoic acid,		0.16	154.6	8.67E-01	1.08	
2333	13.04	375.1297	[M-H]-	1.00	RIBOFLAVIN		0.61	12223.1	4.44E-01	1.13	
2335	12.47	375.1349	[M-H]-	0.17	(-)-Riboflavin; LC-ESI-QT		-0.57	287.7	3.86E-01	1.38	
2829	24.62	412.9666	[M-H]-	1.00	Perfluorooctanoic acid; t		0.25	582.5	8.85E-01	1.07	
3020	23.75	426.9655	[M-H]-	0.83	6:2 Fluorotelomer sulfur		0.48	1551.0	4.68E-01	1.43	
3079	18.07	431.2114	[M-H]-	1.00	5-hydroxy-2,2,6,6-tetran		-0.33	1927.8	5.77E-01	1.29	
3087	16.59	432.2042	[M+FA+H]-	1.00	Guan-fu base Y		-0.54	534.6	1.69E-01	1.67	
3235	16.34	445.0759	[M-H]-	0.50	apigenin-7-O-glucuron		0.80	4377.8	2.65E-01	166.64	
3236	15.56	445.0761	[M-H]-	0.83	apigenin-7-O-glucuron		1.00	28256.1	8.87E-03	164.09	
3387	19.72	459.0892	[M-H]-	0.50	oroxindin		0.79	955.0	2.90E-01	43.35	
4538	12.56	621.1078	[M-H]-	0.50	4'-O-GlcA-7-O-GlcA Api		0.82	3665.6	2.37E-01	130.34	

All the ions in the table can be highlighted, copied and transferred to Excel

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	A	B	C	D	E	F	G	H	I	J	K
1	ID	RT	Mass	Ion	Fraction		Correl	S/N	p-value	FC	
2	20	26.03	96.96125	[M-H]-	1	Phosphoric acid	-0.46775	70.35927	0.870505	1.046085	
3	21	17.95	96.96135	[M-H]-	0.333333	Phosphoric acid	-0.22581	11.09236	0.5714	1.266831	
4	44	14.42	121.0303	[M-H]-	1	2-Hydroxybenzaldehyde	-0.02084	230.3153	0.392505	1.58231	
5	56	7.76	129.0212	[M-H]-	1	CITRACONIC ACID	0.207015	89.03239	0.344172	1.530682	
6	57	8.26	129.0212	[M-H]-	0.333333	CITRACONIC ACID	0.435113	34.15426	0.218116	1.594494	
7	63	10.07	131.0361	[M-H]-	0.333333	Glutaric acid; LC-ESI-QTOF; MS2; CE	0.611154	81.5596	0.644436	1.588785	
8	84	12.12	144.0458	[M-H]-	1	4-Hydroxyquinoline	0.968062	268.3839	0.609065	1.246376	
9	85	17.14	144.0478	[M-H]-	1	4-Hydroxyquinoline	0.470719	13043.28	0.380413	1.298935	
10	86	18.64	144.048	[M-H]-	0.5	4-Hydroxyquinoline	0.285642	118.1601	0.333711	1.610268	
11	95	17.92	148.0242	[M-H]-	1	Benzyl Isothiocyanate	0.087637	150.4269	0.67476	1.374674	
12	96	13.21	149.0088	[M-H]-	0.166667	L-(+)-tartaric acid; LC-ESI-QTOF; MS2; CE	-0.42978	19.05287	0.4837	1.8518	
13	97	25.66	149.0106	[M-H]-	0.333333	(R,R)-TARTARIC ACID	-0.43203	52.50727	0.54429	1.590465	

Clean up the columns to make it easier to appreciate the data

	ID	RT	Mass	Ion	Fraction		Correl	S/N	p-value	FC
2	20	26.03	96.9613	[M-H]-	1.00	Phosphoric acid	-0.4678	70.36	0.87050	1.05
3	21	17.95	96.9614	[M-H]-	0.33	Phosphoric acid	-0.2258	11.09	0.57140	1.27
4	44	14.42	121.0303	[M-H]-	1.00	2-Hydroxybenzaldehyde	-0.0208	230.32	0.39251	1.58
5	56	7.76	129.0212	[M-H]-	1.00	CITRACONIC ACID	0.2070	89.03	0.34417	1.53
6	57	8.26	129.0212	[M-H]-	0.33	CITRACONIC ACID	0.4351	34.15	0.21812	1.59
7	63	10.07	131.0361	[M-H]-	0.33	Glutaric acid; LC-ESI-QTOF; MS2; CE	0.6112	81.56	0.64444	1.59
8	84	12.12	144.0458	[M-H]-	1.00	4-Hydroxyquinoline	0.9681	268.38	0.60907	1.25
9	85	17.14	144.0478	[M-H]-	1.00	4-Hydroxyquinoline	0.4707	13043.28	0.38041	1.30
10	86	18.64	144.0480	[M-H]-	0.50	4-Hydroxyquinoline	0.2856	118.16	0.33371	1.61
11	95	17.92	148.0242	[M-H]-	1.00	Benzyl Isothiocyanate	0.0876	150.43	0.67476	1.37
12	96	13.21	149.0088	[M-H]-	0.17	L-(+)-tartaric acid; LC-ESI-QTOF; MS2; CE	-0.4298	19.05	0.48370	1.85
13	97	25.66	149.0106	[M-H]-	0.33	(R,R)-TARTARIC ACID	-0.4320	52.51	0.54429	1.59

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Reorganizing the data according to RT

	RT	Mass	Ion	Fraction		Correl	S/N	p-value	FC
258	6.51	191.0212	[M-H]-	0.67	CITRATE	0.0265	59.66	0.51275	1.29
677	6.51	243.0623	[M-H]-	0.83	Pseudouridine	0.6598	57.52	0.34761	1.37
678	7.37	243.0630	[M-H]-	0.50	Pseudouridine	0.4058	48.79	0.38523	1.53
259	7.37	191.0213	[M-H]-	0.17	Citric acid	-0.0381	25.39	0.63893	1.23
56	7.76	129.0212	[M-H]-	1.00	CITRACONIC ACID	0.2070	89.03	0.34417	1.53
167	7.76	173.0114	[M-H]-	1.00	cis-Aconitate	0.2025	231.84	0.37510	1.51
57	8.26	129.0212	[M-H]-	0.33	CITRACONIC ACID	0.4351	34.15	0.21812	1.59
1151	8.81	283.0680	[M-H]-	0.50	Xanthosine; LC-ESI-QTOF; MS2; CE	0.4278	74.18	0.49296	1.53
1356	8.93	296.1021	[M-H]-	1.00	N2-Methylguanosine	0.5310	330.68	0.49288	1.36
1565	9.83	310.1140	[M-H]-	1.00	N2,N2-Dimethylguanosine	0.2243	264.50	0.51888	1.36
460	9.96	218.1030	[M-H]-	0.83	D-PANTOTHENIC ACID	-0.5419	603.07	0.49295	1.91
63	10.07	131.0361	[M-H]-	0.33	Glutaric acid; LC-ESI-QTOF; MS2; CE	0.6112	81.56	0.64444	1.59
114	11.44	157.0404	[M-H]-	0.33	ALLANTOIN	0.5857	23.50	0.41526	1.42

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Organizing the data according to p-value

RT	Mass	Ion	Fraction		Correl	S/N	p-value	FC
15.57	269.0468	[M-H]-	0.50	Emodin	-0.5117	2344.65	0.00537	81.25
15.56	445.0761	[M-H]-	0.83	apigenin-7-O-glucuronide	-0.5247	28256.12	0.00887	164.09
16.89	167.1055	[M-H]-	0.83	Chrysanthemic Acid	0.3701	115.90	0.01392	1.35
16.07	204.0672	[M-H]-	1.00	Indolelactic acid	0.1934	5353.15	0.02399	1.41
16.07	158.0597	[M-H]-	1.00	Indole-3-acetaldehyde; LC-ESI-QTOF; MS2; CE	0.1690	110.97	0.09263	1.49
17.62	174.0565	[M-H]-	1.00	Indoleacetic acid; LC-ESI-QTOF; MS2; CE	0.4052	2283.56	0.11465	1.92
21.80	311.1691	[M-H]-	1.00	Triptophenolide	-0.7362	2340.87	0.14646	1.16
14.14	193.0354	[M-H]-	0.67	Glucuronate	0.8812	134.13	0.16489	1.42
22.60	311.1691	[M-H]-	1.00	Triptophenolide	-0.9200	1109.05	0.16520	1.14
15.21	165.0575	[M-H]-	0.33	3-(3-Hydroxyphenyl)propionic acid	0.0486	235.63	0.17098	12.25
19.69	325.1794	[M-H]-	0.17	Dodecylbenzenesulfonic acid	-0.5846	427.25	0.17189	1.26
8.26	129.0212	[M-H]-	0.33	CITRACONIC ACID	0.4351	34.15	0.21812	1.59
18.02	269.0439	[M-H]-	1.00	Apigenin; LC-ESI-QTOF; MS2; CE	-0.4217	7616.71	0.22022	11.24
12.56	621.1078	[M-H]-	0.50	4'-O-GlcA-7-O-GlcA Apigenin (NMR)	-0.4362	3665.59	0.23690	130.34
16.89	204.0664	[M-H]-	1.00	N-Cinnamoylglycine	0.4430	2884.94	0.26166	2.43
21.40	193.0399	[M-H]-	1.00	D-(+)-Galacturonic acid	0.7855	412.51	0.26947	1.50
18.02	432.2045	[M+FA-H]-	0.33	Guan-fu base Y	0.7312	615.06	0.26950	1.52
16.95	187.0982	[M-H]-	1.00	Azelaic acid (Not validated); PlaSMA ID-221	0.5178	3418.23	0.27544	1.31
20.53	269.0465	[M-H]-	0.83	Aloe-emodin	-0.4072	2886.64	0.28086	38.19
19.72	459.0892	[M-H]-	0.50	oroxindin	-0.4125	955.03	0.28982	43.35
18.64	144.048	[M-H]-	0.50	4-Hydroxyquinoline	0.2856	118.16	0.33371	1.61
25.48	157.039	[M-H]-	0.50	ALLANTOIN	0.5537	20.31	0.34204	1.63
7.76	129.0212	[M-H]-	1.00	CITRACONIC ACID	0.2070	89.03	0.34417	1.53
25.69	325.1855	[M-H]-	1.00	Dodecylbenzenesulfonic acid	0.9645	3118.95	0.34721	1.31
6.51	243.0623	[M-H]-	0.83	Pseudouridine	0.6598	57.52	0.34761	1.37
11.45	181.0501	[M-H]-	0.83	DL-3-(4-Hydroxyphenyl)lactic acid; LC-ESI-QTOF; MS2; CE	-0.1964	85.72	0.35649	1.50
24.04	265.1473	[M-H]-	1.00	C12-AS (TENTATIVE)	-0.4008	1845.40	0.35770	2.09
14.54	191.0211	[M-H]-	0.33	CITRATE	0.5951	40.59	0.37175	1.74

These are probably
genistein metabolites

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More information about the experiment

- Although you might think that if you take mice on controlled diet with and without genistein, the variation in observed metabolites would be smaller
- In the experiment described in the [PLoS One paper](#), what we had done is to collect feces from patients with advanced breast cancer who were undergoing chemotherapy
- The fecal organisms were introduced into germ-free mice to establish a close-to-humanized microbiome in each mouse
- **Therefore, the mice had individualized, humanized microbiomes and therefore were not uniform**

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Multivariate statistics

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Set up for PCA analysis

PCA setting

Maximum principal component: **Set this**

Scale method: **Set this**

Transform method:

Metabolite selection

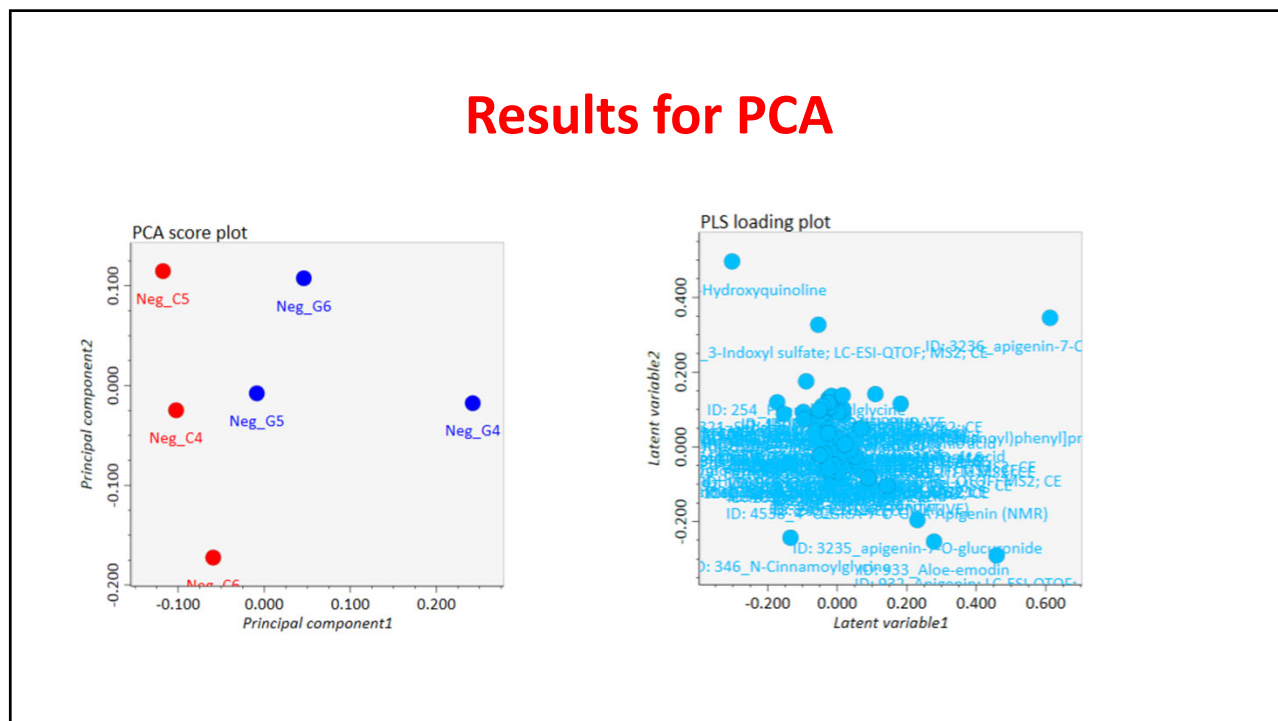
MS2 matched MS1 matedched Unknown

Note – selecting the known compounds

Done

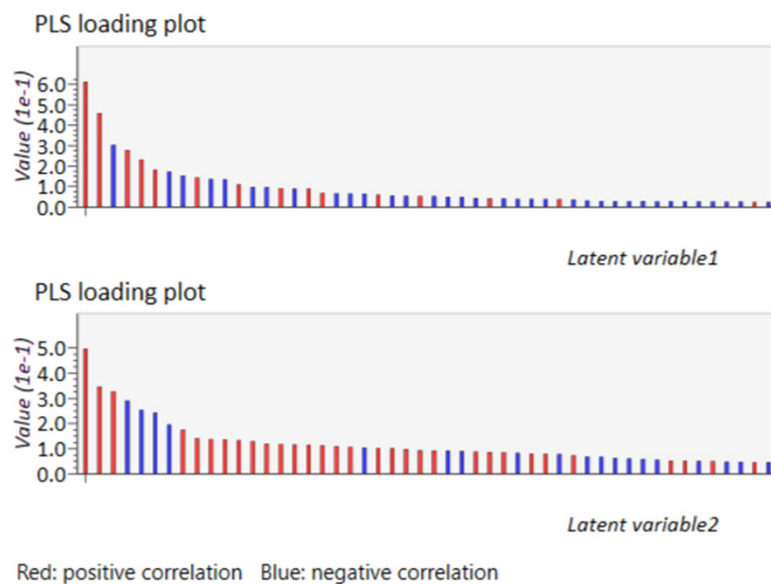
27

Results for PCA



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Plot of the components of the PCA



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Set up for PLSDA analysis

PLS setting

General setting

Auto fit

Scale method: Pareto scale

Transform method: None

Components number: 2

PLS method

PLSDA PLSR OPLSDA OPLSR

Metabolite selection

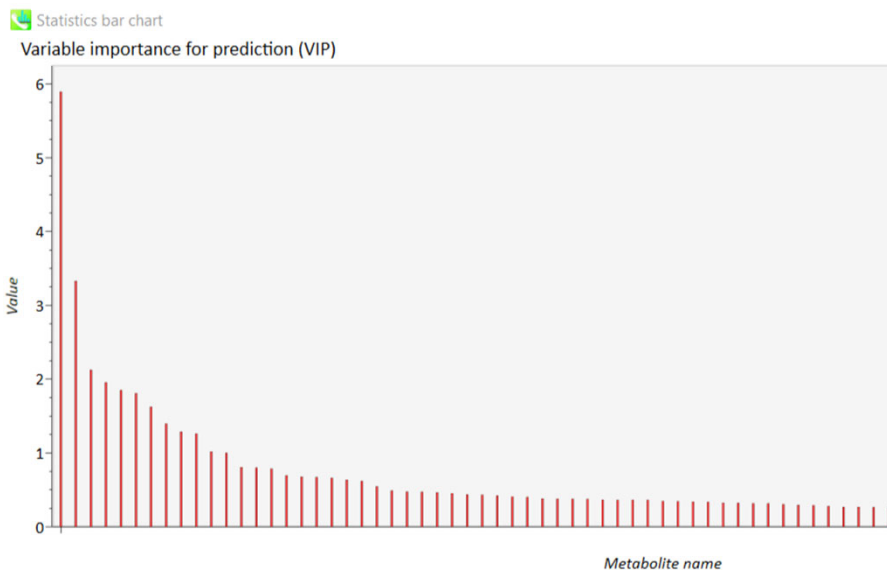
MS2 matched MS1 matched Unknown

- (1) Set Y (response) variables at menu->option->file property.
 (2) For (O)PLS-DA, use a binary (0 or 1) value as the response.
 * Non-zero values are recognized as 1 in (O)PLS-DA testing.
 (3) For (O)PLS-R, set sequential values.

Done

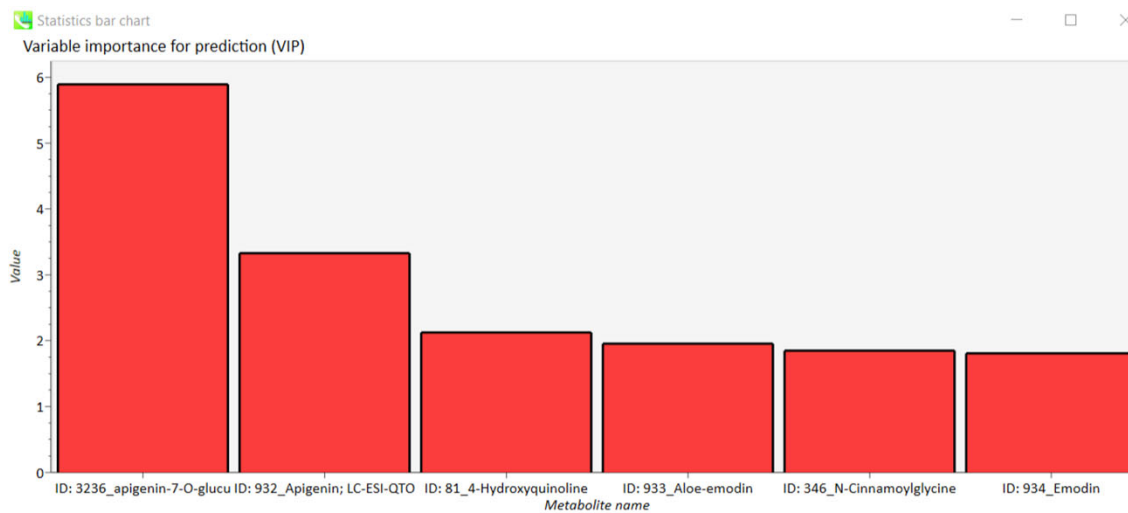
30

PLSDA VIP plot



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Expanded PLSDA VIP showing the top metabolites



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PLS setting

General setting

Auto fit

Scale method: Pareto scale

Transform method: None

Components number: 2

PLS method

PLSDA PLSR OPLSDA OPLSR

Metabolite selection

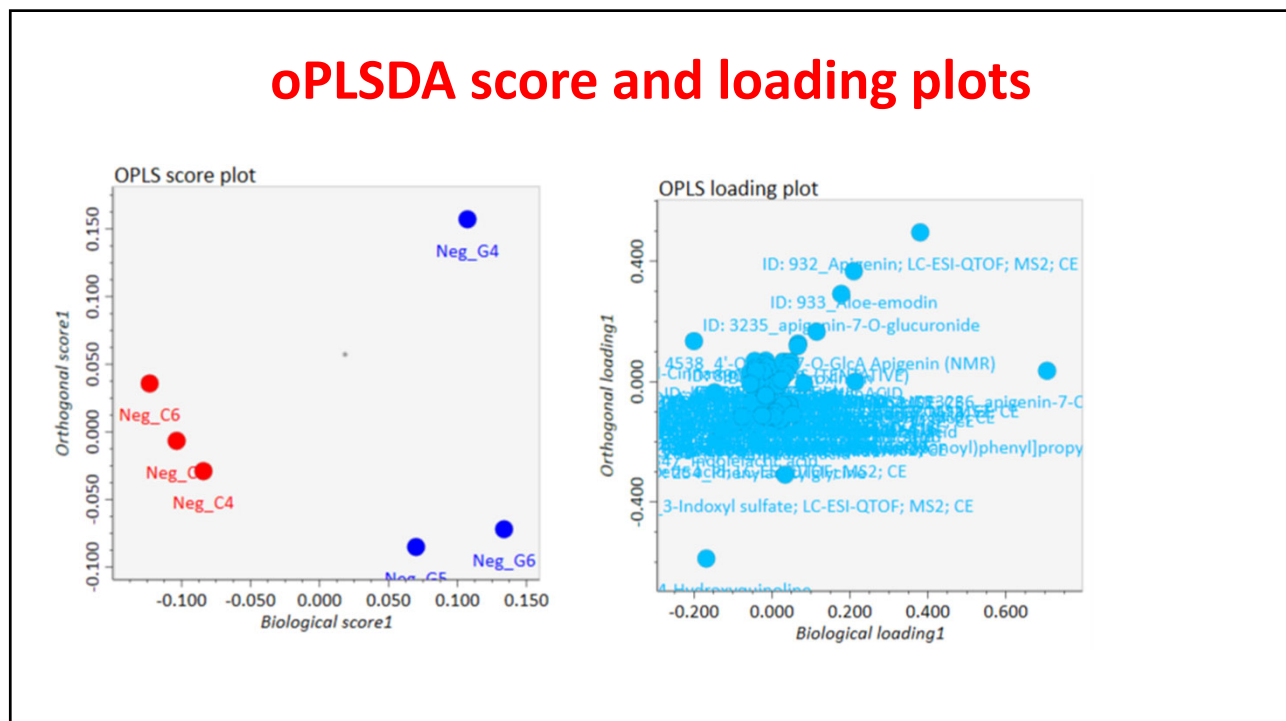
MS2 matched MS1 matched Unknown

(1) Set Y (response) variables at menu->option->file property.
(2) For (O)PLS-DA, use a binary (0 or 1) value as the response.
* Non-zero values are recognized as 1 in (O)PLS-DA testing.
(3) For (O)PLS-R, set sequential values.

Done

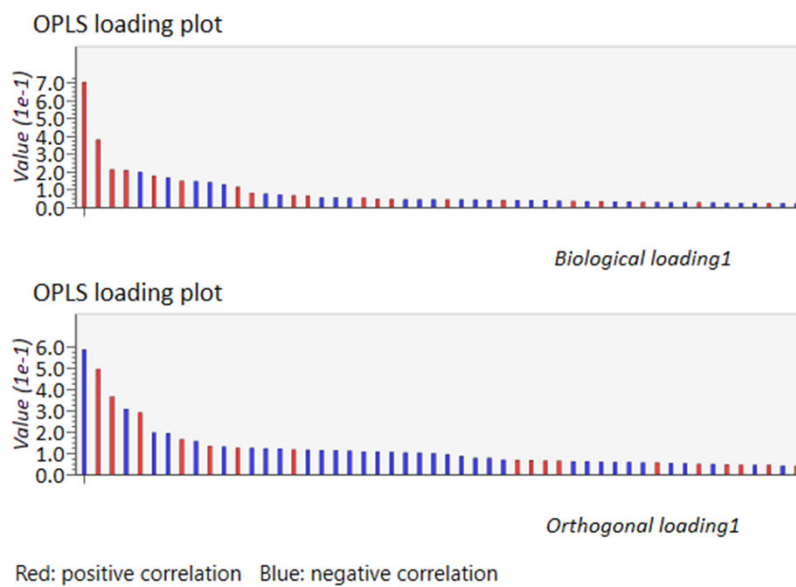
Setting up the oPLSDA analysis

33



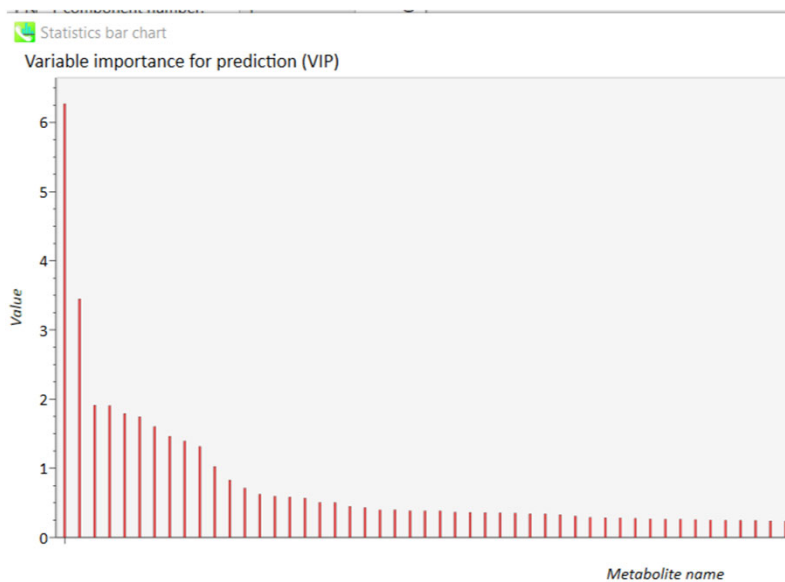
34

oPLSDA component plots



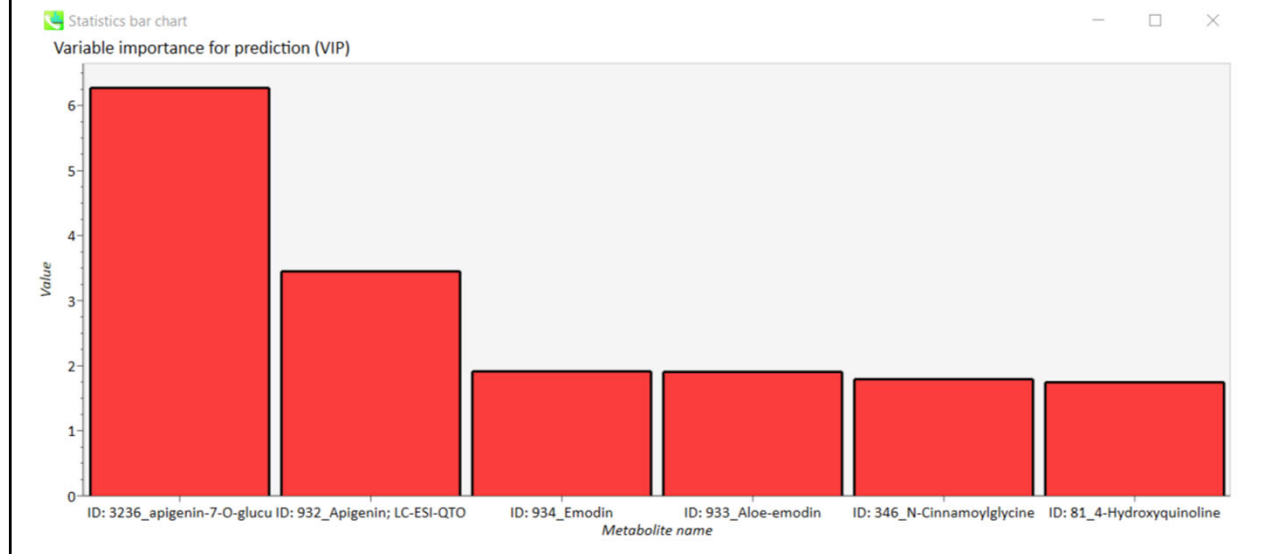
35

oPLSDA VIP components

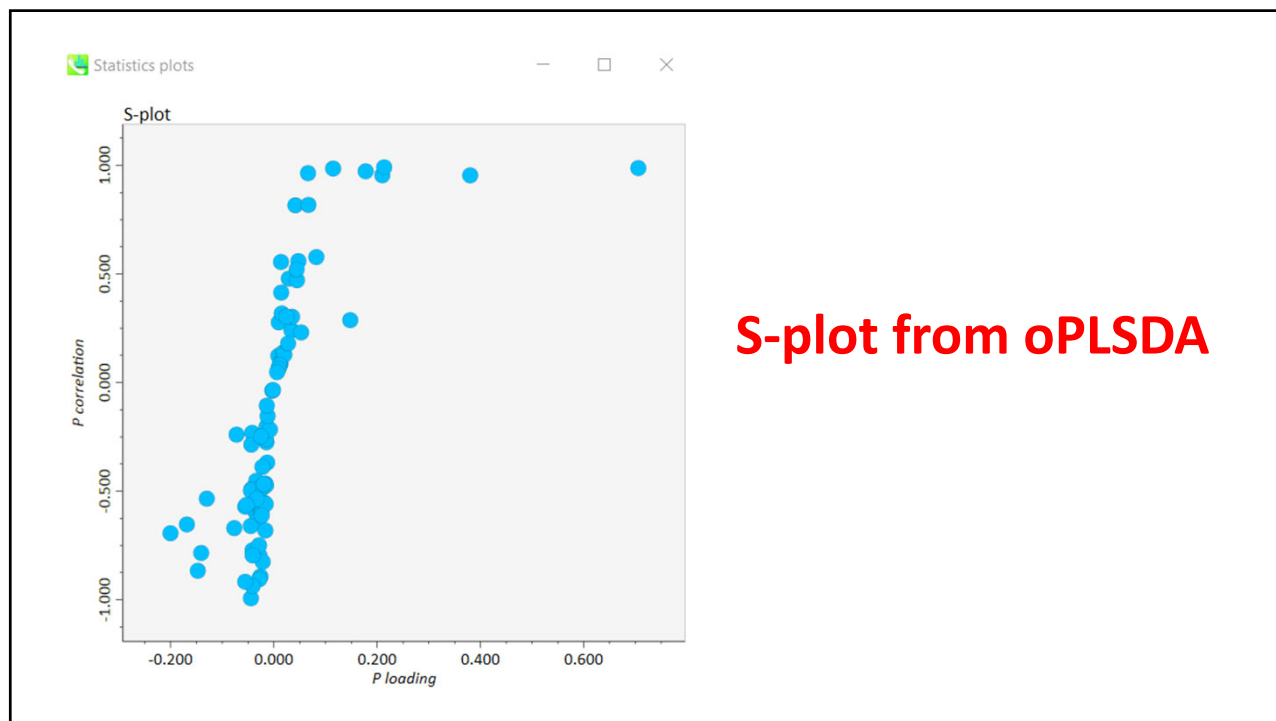


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Expanded oPLSDA VIP items



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Alignment result export

Directory: C:\Users\sbarne\Desktop\Class 2020\class 2-10-20 fi

Export option

File: alignmentResult_2020_2_11_7_39_25

Raw data matrix (Height) Peak ID matrix
 Normalized data matrix Retention time matrix
 Raw data matrix (Area) m/z matrix
 *Export as mztab-M MS/MS included matrix
 GNPS export
 S/N matrix export
 Representative spectra
 Parameter

Filtered by blank peaks (must be checked in alignment parameter setting)

Filtering by the ion abundances of blank samples

Missing value option

Replace zero values with 1/10 of minimum peak height over all samples

Isotope labeled tracking option Target file

Filtering by the result of isotope labeled tracking Neg_G6

Export format: mgf Spectra type: centroid

This outputs information about normalized areas of the metabolite peaks in each sample

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Alignment result export

Directory: C:\Users\sbarne\Desktop\Class 2020\class 2-10-20 fi

Export option

File: alignmentResult_2020_2_11_7_39_25

Raw data matrix (Height) Peak ID matrix
 Normalized data matrix Retention time matrix
 Raw data matrix (Area) m/z matrix
 *Export as mztab-M MS/MS included matrix
 GNPS export
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 Representative spectra
 Parameter

Filtered by blank peaks (must be checked in alignment parameter setting)

Filtering by the ion abundances of blank samples

Missing value option

Replace zero values with 1/10 of minimum peak height over all samples

Isotope labeled tracking option Target file

Filtering by the result of isotope labeled tracking Neg_G6

Export format: mgf Spectra type: centroid

This output file allows us to look at adduct ions that are related to the known metabolite ions

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